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# A Review: Analytical methods for heavy metals determination in environment and human samples

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## **ABSTRACT**

Heavy metals are vital and necessary in our daily lives. Moreover, if the amounts of heavy metals are more than the acceptable amounts (mentioned by WHO) in soil, water, and air, indeed, they cause a lot of diseases in human bodies. Therefore, monitoring and measuring the amounts of heavy metals which are arduous and difficult are so important. In this review paper, a lot of studies which have been carried out on the determination and quantification of heavy metals in human bodies, soil, and water are considered. Moreover, the effect of toxicity of each heavy metal on human health is assessed. According to WHO, EPA, NIOSH, ACGIH, and clinical chemistry, determination of heavy metals such as Cd, Pb, Zn, Hg, Cu, Mn is very important in human body and Environmental matrixes. For example, the range concentration of heavy metals in human body such as, cadmium is about 0.3 to 0.98, and for cooper, it is about 0.01 - 1.107 ppm. In addition, most obtained results demonstrate that the lowest and highest range concentration of heavy metals in blood of human body has been equal to 0.56-8.78 ppm and 0.08 - 4.67 ppm respectively. Finally, in this review paper, the approaches of quantification of heavy metals using atomic absorption spectrometry, gas chromatography, inductively coupled plasma mass spectrometry etc. in human bodies, water, and soil are assessed.

## 1. Introduction

Many different techniques have been applied for determination of heavy metal in different samples. Atomic absorption spectrometry (AAS) is a quantitative method of metals analysis which was suitable for the determination of 70 elements by three option accessory (flame, furnace, Hydride generation). AAS measures the concentration of the metals by passing light (wave length emitted) by a radiation source of

\*Corresponding Author: Hamid Shirkhanloo E-mail: Hamidshirkhanloo@yahoo.ca; iranma4@gmail.com https://doi.org/10.24200/amecj.v2.i03.73 a particular element such as Hg, As, Pb and etc. Many metals determined by electrothermal atomic absorption spectrometry (ET-AAS) [1-9], flame atomic absorption spectrometry (F-AAS and FS-AAS) [10-18], atom trapping flame atomic absorption spectrometry (AT-FAAS), Hydride generation atomic absorption spectrometry, Cold vapor-atomic absorption spectrometry (HG-AAS, CV-AAS)[19, 20] and fluorescence spectrometry (XRF) [21-24]. Also, direct current polarography (DCP) [25, 26], differential pulse polarography (DPP), neutron activation analysis (NAA), inductively coupled plasma-optical emission spectrometry (ICP OES) [27, 28], inductively

coupled plasma- mass spectrometry (ICP-MS) [29, 30], UV-VIS [31], HPLC-inductively coupled plasma-mass spectrometry (HPLC - ICP - MS) [32], electrophoresis(EP) [33], Anodic striping voltammetry (AVS) and inductively coupled plasma-mass spectrometry (ICP-MS) was used for heavy metal determination in different matrix [34, 35]. Arida et al showed that the assessment of heavy and toxic metals in the blood and hair by modern analytical techniques in review paper. Heavy metals and toxic metal effect in blood and hair of some smokers was determined by modern analytical techniques. The levels of Hg, Pb, Cd, As, Se, Mn, Zn, Ni, and Cr were determined using inductively coupled plasma-atomic emission spectrometer (ICP-AES) [36]. A review procedure based on heavy metal analyzing in soil was developed with sequential extraction by Zimmerman et al in 2010. They showed that the quantification extraction and determination of heavy contamination in soil cannot give reliable results. They showed, the procedures of community bureau of reference (BCR) and geological society of Canada (GCS) procedures were examined to clarify benefits and limitations of each [37]. Also, determination of heavy metal in blood evaluated with AAS by Batool and Ahmad at 2018. The blood samples were divided into two parts .First for analysis of total blood and other for serum. By centrifuging the blood and serum sample separated. Serum after adding de ionized water analyzed by instrument after digestion. For whole blood samples wet digestion apply before analysis. Before analysis, blood samples were kept in freezer at 4 °C. In this study, wet digestion includes decomposition by acids carried out on hotplate and continued by AAS model Varian AA 240 [38]. Yuan Fu et al used Titanate/TiO<sub>2</sub> as adsorption sorbent for heavy metals determination and adsorption by nanotechnology. The titanate/TiO2 Nanomaterials were synthesized by Biomolecules (lignin). The titanate/TiO2 characterized by XRD, FESEM, TEM, FTIR and BET, and their adsorption for heavy metals were studied. The found the maximum adsorption capacity of heavy metal ions (Pb<sup>2+</sup>, Cu<sup>2+</sup>, Cd<sup>2+</sup>) based on procedure

[39]. Zhang et al showed, the nanomaterials can be treatment in water and wastewater samples. They introduce a novel nanotehecnology which was used by researcher included, the synthesis and physiochemical properties of nanomaterials, carbon based nanomaterial, metal and metal oxides nanoparticles were focused on removal of various heavy metals in water and wastewater Chromium speciation in natural water [40]. samples was developed by solid phase extraction procedure. Cr(VI) can be extracted with ammonium pyrrolidine dithiocarbamate (APDC) pasted on multi walled carbon nanotubes (MWNTs). The total chromium was determined by oxidation of Cr(III) to Cr(VI). The Cr(III) concentration is calculated by subtracting of total chromium and Cr(VI) [41]. Cr (VI) compounds, once inside the bloodstream, are actively transported into red blood cells (RBC) via nonspecific anionic channels and then rapidly reduced to Cr (III) which becomes bound to hemoglobin. As canceroginic effect of Cr (VI) determination of Cr (VI) very important. Also, in Figure 1, metabolism and formation of DNA damage by CrVI at pH 7.4 are shown.

Zendehdel et al studied on a HAp/NaP nanocomposite by adding a synthesized nanohydroxyapatite to zeolite NaP gel. They used HAp/ NaP for the removal of lead(II) and cadmium(II) ions from aqueous solution. This nanocomposite was characterized by FTIR, X-ray, SEM, energydispersive X-ray analysis and thermal analyses [42]. Microwave-assisted method was used for synthesizing of graphene nanosheet/δ-MnO , (GNS/ MnO<sub>2</sub>) composite. TheGNS/MnO<sub>2</sub> as sorbent was used for removal of NI(II) from waters. SEM, TEM and FTIR show that nanoscale  $\delta$ -MnO , deposit well on GNS. The adsorption process was achieved in optimized pH [43]. Ojea-Jiménez et al showed that the removal of mercury from the Ebro River by Colloidal gold nanoparticles (Au NPs). Au NPs have been used for sequestration removal of Hg(II) from multicomponent waters. Under optimized conditions, sodium citrate was reduced Au NPs as the catalyst in the reduction of Hg(II), which is efficiently trapped in the presence of Cu(II) and

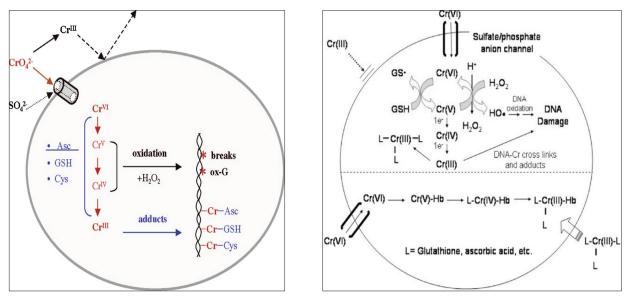


Fig. 1. Metabolism, and formation of DNA damage by Cr (VI) at pH 7.4.

Fe(III) [44]. Many references existed for separation and preconcentration and determination based on LLME, SPE, MSPE, IL/MSPE, USA-MSPE etc. in water, human biological and food samples by nanotechnology [45-56]. Also many developed methods was reported by researchers by green solvents such as ionic liquid for extraction of heavy metals in different matrixes [57-88]. In addition, an advanced approach for separating As(III) and As(V) ions in environmental matrixes was presented by Soylak et al in 2009 [89]. In their research, when dilute KMnO<sub>4</sub> was used to oxidize As(III), then the developed coprecipitation was used to determine total arsenic [89]. The difference between the total arsenic content and As(V) content was calculated as Arsenic(III) [89]. Moreover, in their study, hydride generation atomic absorption spectrometry (HG-AAS) was used to determine arsenic levels. Also, after calculating the factor of preconcentration, it was found out that the factor was equal to 25. In addition, the detection limits (LOD) for As(V) was  $0.012 \mu g. L^{-1}$  [89]. A new approach for solid phase extraction (SPE) of gold(III) by using Dowex M 4195 chelating resin was developed by Soylak et al in 2007 [90]. The quantitative sorption of gold<sup>3+</sup>, effect of flow rates, pH, sorption capacity, eluent types, and the effect of diverse ions on the sorption of Au<sup>3+</sup> were investigated to find optimum experimental conditions [55]. In their work, for

more than 100 cycles of sorption-desorption, the chelating resin was reused without any considerable change in sorption of Au<sup>3+</sup> [90]. In their work, the preconcentration factor was calculated to be 31. Detection limit (LOD) of Au was greater than 1.60  $\mu$ g L<sup>-1</sup> and the recovery values for Au<sup>3+</sup> was greater than 96% [90]. The adsorption capacity of the resin was 8.1 mg.g<sup>-1</sup>. Their proposed approach has been determining Au<sup>3+</sup> in some real soil and water samples were used and showed the best results [90]. For characterizing As(III) biosorption from aqueous solution, green algae (Ulothrix cylindricum) biomass was used by Soylak et al in 2009 [91]. According to their studies, the biosorption capacity of Ulothrix cylindricum biomass was equal to  $67.2 \text{ mg g}^{-1}$  [91]. The outcomes of their work indicated that the processes of biosorption of As<sup>3+</sup> followed well pseudo-second-order kinetics [91]. Separation of Cu<sup>2+</sup> in environmental matrixes by using solid phase extraction on multi-walled CNT (carbon nanotubes) was studied by Soylak and Ercan in 2009 [92]. The optimum adsorption was obtained on MWNTs column in their study when pH was equal to 7.0. The preconcentration factor was equal to 60. Also, the detection limit for  $Cu^{2+}$  was equal to 1.46 µg L<sup>-1</sup> [92]. For atomic absorption spectrometry, separation of trace heavy metals in sediment, urine, and dialysis concentrates by coprecipitation with samarium hydroxide was

carried out by Elci et al in 2003 [93]. In their study, the optimum pH for collecting trace heavy metals on samarium hydroxide was equal to 12.2. Also, coprecipitation occurs after 30 minutes, and when quantitative recovery is governed, and the precision is about 5% for the analytes [93]. Determining heavy Metals in plants, soil, and groundwater which followed high-rate sewage sludge was studied by Speir et al in 2003 [94]. The pH of samples was less than 4. Also, during five years, the concentration of Zn in the soil reduced from 27 mg kg<sup>-1</sup> to 0.04 mg kg<sup>-1</sup>. Moreover, based on their study, there is a direct relation between the concentration of heavy metals in plants and the concentration of heavy metals in the soil [94]. An advanced biological treatment plant for removing heavy metals in groundwater was presented by Scheeren et al in 2007. Groundwater below the zinc production plant was contaminated with sulphate and heavy metals. To avoid contamination of nearby drinking-water aquifers in the future, a treatment plant for extracting contaminated ground water was established by them [95]. Effluent treatment based on activity of Sulphate Reducing Bacteria (SRB) for excess sulphide in solids for achieving the discharge criteria was designed by them. The reducing the sulphate to sulphide and thereby, precipitation of metal sulphides were obtained when anaerobic bacteria are there in the soil or the bacteria are injected [95].

Determination of trace heavy metals in harvested rainwater by ICP-MS was used by Malassa et al in 2018 [96]. Inductively coupled plasma mass spectrometry (ICP-MS) was used to quantify the content of each trace heavy metal (Cr, Co, Mn, Ni, Cu, Zn, Mo, Ag, Cd, Bi, and Pb) in a few rainwater samples. Their results indicated that that the concentrations of Cr, Mn, Ni, Ag, and Pb are more than the WHO limits in drinking water [96]. Contamination of heavy metals in sediments of the Karasu spring in Nigde, Turkey has been investigated by Yalcin and Narin in 2007 [97]. A few sediment matrixes were collected from contaminant sites along the spring. Moreover, concentrations of heavy metals have been determined by X-ray Fluorescence Spectrometer [97]. The amount of Co, Cu, As, Sn, Ni, Zn, Ca, Pb, Al, Fe, Ti, Cr, and Mn contents of the Karasu creek sediments were equal to 18.30-69.00, 12.40-595.0 5.50-345.3, 5.80-15.1, 10.9-64.1, 28.90-103,300, 4.1-356.2, 7.70-37,840, 13,460-109,400, 11,740-62,900, 22.18-59.04, 41.70-369 and 12.09-3,480 mg/kg respectively. Moreover, according to their results, the presence of a contamination in the Karasu creek is obvious. In the case study, all the metal concentrations are more than acceptable limit values. In addition, in the Karasu creek and the Akkaya dam, eutrophication has been being developed. Also, it sounds that the accumulation of heavy metal in the creek is come from the discharge of mine quarries, domestic, and industrial wastes [97]. Separation of arsenic(III) and arsenic(V) by using Streptococcus pyogenes which has been immobilized on Sepabeads SP 70 resin has been considered by the approach of solidphase extraction by Uluozlu et al in 2010 [98]. In sample solutions, hydride generation atomic absorption spectrometry (HGAAS) has been used to quantify the arsenic levels [98]. Their procedure has been governed when recovery of As<sup>3+</sup> was more than 95%. Moreover, recovery of As<sup>5+</sup> which was obtained was less than 5%. By using KI and ascorbic acid, As(V) has been reduced; in addition, after 1 hour later, for determining total arsenic, their system has been applied. As<sup>5+</sup> has been calculated by subtracting the total As from As(III) content [98,99]. Some experimental parameters such as pH, amount of microorganism, sample volume have been assessed in their study. The capacity of biosorbent for  $As^{3+}$  has been equal to 7.3 mg g<sup>-1</sup>. The preconcentration factor has been found to be equal to 36 [99]. The relative standard deviation has been obtained less than 8% [99]. Moreover, the contamination of trace heavy metal ions in drinking water samples has been investigated by Tuzen et al in 2005. According to their studies, if the amount of lead in drinking water samples is more than 10 microgram per liter, brain tissues and kidneys will be damaged [100]. Assessment of the amount of each metal in Drinking Waters from the

Tokat-Black Sea Region of Turkey was studied by Tuzen and Soylak in 2006. In their study, atomic absorption spectrometry was used to determine the amount of Cr, Ni, Cu, Mn, Zn, Fe, Co, and Al ions. According to their study, the amount of the heavy metals was lower than the maximum acceptable limits which were set by the World Health Organization [101-138]. In this review paper, the amounts of heavy metals and their toxicity in water, soil, air, and human bodies which have been studied and quantified by other researchers during the last decade are considered.

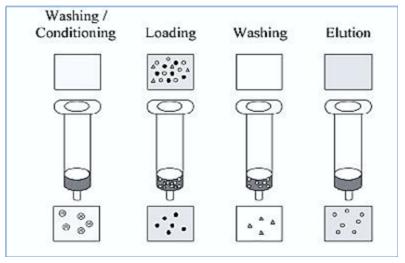
## 2. Experimental

Kaipei Qiu used ultrafine mesoporous magnetite nanoparticles (UFMNPs) for removal of heavy metals from waters (Scheme 1). Ultrafine magnetite NPs(UFMNPs, Fe<sub>3</sub>O<sub>4</sub>) were synthesized by coprecipitation method and successfully used as adsorbents for the removal of Pb<sup>2+</sup>, Cd<sup>2+</sup>, Cu<sup>2+</sup>, and Ni<sup>2+</sup> ions from contaminated river water. Also, the adsorption/desorption tests demonstrated that the ultrafine Fe<sub>3</sub>O<sub>4</sub> NPs were reusable and can be used several times [139].

Gollavelli et al and Luo et al used Magnetic Graphene and magnetic chitosan/cellulose microspheres for removal of heavy metal from dinking waters, waters [140,141]. Mahmoudi ,Chen and Neyaz can be extracted heavy metals from different matrix by iron oxide nanoparticles (SPIONs),  $Fe_3O_4$  @ SiO<sub>2</sub> and functionalized iron oxide nanomaterials [142-144]. Tuzen et al showed a novel method based on switchable hydrophilicity microextraction syringe system (µS-SHS) by electrothermal atomic absorption spectroscopy (ETAAS). The proposed method was developed for separation and determination of vanadium in waters by complexing 2-(5-Bromo-2-pyridylazo)-5-diethylaminophenol (5- Br-PADAP). In this study, LOD, EF, and RSD were achieved 7.5 ng L-1, 120 and 2.91 %, respectively. In addition, the µS-SHS is consisting of micropipette tip and syringe system was designed. First, the 10 mL of real sample was placed on beaker and then, 5- Br-PADAP solutions as ligand were used by sucking into syringe system. Finally, 0.1 mL of decanoic acid as a hydrophobic effect sucked into to solution which was followed by 500 µL of 1.0 mol L-1 Na<sub>2</sub>CO<sub>2</sub> solution as a switchable hydrophobicity for the decanoic acid (CH<sub>3</sub>(CH<sub>2</sub>)<sub>8</sub>COOH) in liquid phase. The surface response methodology was used to calculate response surface between [pH]/[V of SHS] and [pH]/[V of Na<sub>2</sub>CO<sub>3</sub>] of the vanadium. The results showed, high recovery based on 112 µL of SHS and 624 µL of Na<sub>2</sub>CO<sub>3</sub> was obtained at pH=3.28. The design of experiments (DOE) was used for analyzing data and optimizing by surface response methodology [102]. Xiaogang Luo et al explained a novel sorbent for absorption heavy metals from waters. The carboxyl decorated magnetite ( $Fe_3O_4$ ) nanoparticles (MN-CA) were

obtained by modifying the magnetite surface with

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Scheme 1. SPE method for extraction heavy metals from liquids

citric acid. In this research, the combination of specifically modified magnetic nanoparticles and activated carbon(AC) to cellulose may create new nanocomposite adsorbents, which possess high adsorption capacity for heavy metals removal and easy recovery by magnetic field. It could be deduced that electrostatic attraction between negatively charged magnetic cellulose-based beads (MCB) samples and positively charged Cu<sup>2+</sup>, Pb<sup>2+</sup>, and Zn<sup>2+</sup> was the initial driving force of heavy metals binding to the adsorption site of the adsorbents [145]. Researcher such as Shannon, Luo, Gericke, Karami and Zhao et al worked for removal heavy metals from water, water Purification, wastewater treatment and methyl orange removal by novel technique with different sorbents such as; activated cellulose Microspheres, magnetic functional magnetite nanoparticles, and cellulose beads, MWCNTs/Fe<sup>3</sup>O<sub>4</sub>/PANI hierarchical magnetic composite [146-150]. For determining the amount of metals in drinking water samples, the Perkin Elmer Analyst 700 atomic absorption spectrometer was used by Tuzen and Soylak in 2006 [101]. A 10 cm long slot-burner head, a lamp and an airacetylene flame were applied. In solid phase extraction (SPE) studies, a glass column with 100 mm high and 10 mm in diameter was applied [101]. The solid phase materials such as Amberlite XAD-1180, Amberlite XAD-4 and Diaion HP-2MG was purchased, and they were prepared with the washing steps as reported previously in literature. They collected the drinking water samples in prewashed (with detergent, doubly de-ionized distilled water, diluted HNO<sub>3</sub> and doubly de-ionized distilled water, respectively) polyethylene bottles from eight stations in Tokat and villages around Tokat in July 2004. The samples were obtained from the water pump directly after the water had run for at least twenty minutes. They filtered the samples by using a Millipore cellulose membrane with a 0.45 µm pore diameters [101]. After the process of filtration, nitric acid was used to acidify the samples to 1%, and then the samples were stored in 1 L polyethylene bottles. The samples were thereby stored at 4 °C for a short time, afterwards they were analyzed until the

changes of the physicochemical form of the metals were minimized [101]. Electroplating industry, tanneries, electronics manufacturing industry, coalfired power plants and mining operation are key sources of heavy metal pollution in water [151]. Lead (Pb) is discharged into water sources from various industries and is highly toxic to human beings even at trace concentrations. Maximum contaminant level of Pb(II) in drinking water is set at 15 µg L-1 by US Environmental Potential Agency (US EPA). Chromium(Cr), another toxic heavy metal pollutant, is mainly present in the effluents of leather tanning, electroplating and chromate preparations. Chromium is found in two oxidation states Cr(III,VI) and Cr (VI) is more toxic. World Health Organization (WHO) guideline for Cr(VI) in drinking water is 50 µg L<sup>-1</sup>. So determination and separation this pollutant from waters with novel technique based on sorbent adsorption is very important [151]. Santhosh et al showed lead and Cr simply extracted from water solutions by PG-C and CNF-C [151]. By using thermal process, cobalt ferrites (CoFe<sub>2</sub>O<sub>4</sub>) paste on porous graphene (PG) and carbon nanofibers (CNF)), which was named PG-C and CNF-C nanocomposites as magnetic adsorbents [151]. In Figure 2, removal of heavy metals using nanographene is shown.

Sarika Singh et al used a functional oxide nanomaterials and nanocomposites for the removal of heavy metals and dyes in liquid phase. They showed that the oxide-based nanomaterials, such as Fe<sub>3</sub>O<sub>4</sub>, ZnO and TiO<sub>2</sub> for the removal of different toxic metals and organic dyes at optimized pH. Magnetic nanoparticles as excellent sorbent can be used as highly effective, efficient and economicallyviable for removal toxic metals with advantages of easy separation under a magnetic field for reuse. Shen et al have observed that the adsorption efficiency of Ni2+, Cu2+, Cd2+ and Cr6+ ions by Fe<sub>3</sub>O<sub>4</sub> nanoparticles is strongly dependent on pH, temperature, and the amount of the adsorbent. In optimized conditions, 3.5 mg mL<sup>-1</sup> dose of nanoadsorbent with an optimum pH=4 was used for removal heavy metals from contaminated water [152]. Phoebe Zito Ray et al studied on Inorganic

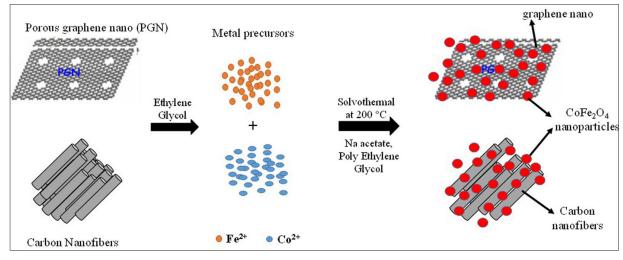


Fig. 2. Removal of heavy metals using nano-graphene [151].

nano-adsorbents for the removal of heavy metals and arsenic in aqueous solutions. They were explained that metal oxides and CNTs as nanoadsorbents were used for heavy metal removal in aqueous solutions. Phoebe Zito Ray et al used the iron oxide (hematite, magnetite and maghemite), carbon nanotubes (CNT), and metal oxide based (Ti, Zn) and polymeric nanoadsorbents for this study. Based on reults, metal oxides can have increased surface area and possess favorable sorption to heavy metals. ZnO nanoplates showed complete removal of Cu2+ in aqueous solutions [153]. Recently, CNTs and nZVI synthetized by coupling these nanomaterials and was efficient used for removal of Cr(VI), Se and Co from polluted water by Giorgio Vilardi et al. Regeneration studies were performed fixing CNTs-nZVI concentration and the initial metal concentration equal to 10 mg L<sup>-1</sup>, as already reported in a previous study [154]. Experimental results of adsorption batch tests showed that the removal efficiency of nanomaterials was in decreasing order: CNTnZVI>nZVI>CNTs>CNTs-nZVI, except for the Cr(VI) that nZVI was the most efficient material. Furthermore, the removal of Cr(VI) was found to not be affected by varying the pH in the case of using nZVI, but decreased with increasing pH when using the other three nanomaterials [154]. Also, Majed Alrobaian and Hassan Arida studied on Assessment of Heavy and Toxic Metals in the Blood and Hair of Saudi Arabia Smokers Using Modern Analytical Techniques. The levels of some

selected heavy and toxic metals (e.g.; Hg, Pb, Cd, As, Se, Mn, Zn, Ni, and Cr) were determined using inductively coupled plasma-atomic emission spectrometer (ICP-AES). Prior to the analysis, the blood and hair samples of Saudi Arabia smokers were collected, treated, and digested by microwave digestion system. The validation measurement of ICP-AES was performed using working calibration solutions of the investigated toxic heavy metal ions (Hg, Pb, Cd, As, Se, Mn, Zn, Ni, and Cr) [155]. Kaiser, Zheng and Feng was introduced a novel two-dimensional polymers synthesis with nano size in water which was different application such as heavy metal removal. They reported the controlled synthesis of few-layer two-dimensional polyimide crystals on the surface of water through reaction between amine and anhydride monomers, assisted by surfactant monolayers [156]. Heavy metals such a lead or cadmium has devastating health effects on human body. So, removal of heavy metals from industrial wastewater and drinking water is very important. Rosillo-Lopez et al showed that carboxylated graphene nanoflakes (cx-GNF) and nano-graphene oxide (nGO) can be extracted  $Fe^{2+}$ , Cu<sup>2+</sup>, Fe<sup>3+</sup>, Cd<sup>2+</sup> and Pb<sup>2+</sup> ions from water sample. The sorption capacity for  $Pb^{2+}$ , , is more than six times greater for the cx-GNF compared to GO which is attributed to the efficient formation of lead carboxylates as well as strong cation– $\pi$  interactions [157]. Zhao et al introduced a novel multithroughput dynamic microwave-assisted leaching

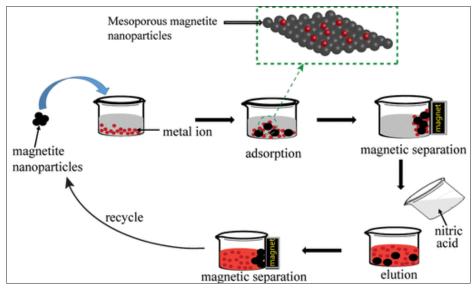
(MDMAL) combined with ICP-AES was used for the simultaneous determination of Cu, Mn, Zn and Pb in soil. The MDMAL method was more rapid than the simple digestion method or microwave digestion process and could be an alternative approach for the analysis of heavy metals in solid samples [158]. Aaron et al reported an efficient chemical strategy for the crosslinking of polymer chains based on the N- and C- of proteins(CPC $\rightarrow$ N/C-P), producing hybrid hydrogels which was related to biomolecules. This compound is largely independent of the protein's sequence; it can in principle be used to incorporate of polypeptide into a polymer based on uniform linkages. They used cysteine for this strategy to convert metallothioneins into a recyclable polymer gel that can remove heavy metals from water samples [159]. Shirkhanloo et al reported a novel method based on amine-functionalized mesoporous silica UVM-7 as nanoadsorbent for manganese (II) and manganese (VII) extraction in waters by ultrasound assisted-dispersive-micro-solid phase extraction (US-D-µ-SPE) procedure. They were used NH2-UVM7 for speciation of Mn(II) and Mn(VII) ions at pH of 8 and 5, respectively which was determined by(AT-F AAS. The validation of the method was also performed by the standard reference material [160]. Also, Shirkhanloo et al introduced a carboxyl-functionalized nanoporous graphene (NG-COOH) as solid phase sorbent for speciation of trace Hg (II) and R-Hg (CH<sub>2</sub>Hg<sup>+</sup> and  $C_2H_sHg^+$ ) in waters by US-D-IL-  $\mu$ -SPE (pH=8). Ionic liquid was used for trapping of sorbent from water sample. After back extraction, Hg (II) concentration determined with flow injectioncold vapor-atomic absorption spectrometry (FI-CV-AAS) [161]. Also they were reported a new method based on ultrasound assisted-dispersive solid-liquid multiple phase microextraction (USA-DSLMPME) for speciation of trace amounts of inorganic arsenic (III and V) and total organic arsenic (TOA) in waters and human urine samples. By procedure NH2-UVM7 were immersed in ionic liquid (IL) as an extraction phase. Then, the mixture of NH2-UVM7 and IL/acetone [HMIM] [PF6]/AC) was injected to sample at pH of 3.5.

After ultrasonic, As(V) anion extracted by binding to amine group. Total amount of inorganic arsenic (TIA) was determined after oxidization of As(III)to As(V) by ETAAS and As(III) was calculated by the subtracting TIA and As(V) content [162].

## 3. Results and Conclusions

Kaipei Qiu was introduced a novel adsorption process for heavy metals from waters. In the adsorption process, the desorption and repeatability are significant parameters for developing new adsorbents for practical applications. The heavy metals such as, Pb2+, Cd2+, Cu2+, and Ni2+ were adsorbed by UFMNPs which was shown in Scheme 2. The adsorption kinetics is one of the important characteristics defining the effectiveness of an adsorbent, which describes the solute uptake rate by controlling the diffusion process and the residence time of an adsorbate uptake at the solid/solution interface. The results in this quaternary Pb-Cd-Cu-Ni system indicated that the rate constant of  $Pb^{2+}$  (0.045 g mg<sup>-1</sup> min<sup>-1</sup>) was higher than the rate constants of Cu<sup>2+</sup> (0.008 g mg<sup>-1</sup> min<sup>-1</sup>), Cd<sup>2+</sup> (0.005  $g mg^{-1} min^{-1}$ ), and Ni<sup>2+</sup> (0.003 g mg<sup>-1</sup> min<sup>-1</sup>), further demonstrating the strong affinity of Fe<sub>3</sub>O<sub>4</sub> NPs for Pb<sup>2+</sup>[139].

Soylak and Tuzen studied on the amounts of Cr, Cu, Ni, Zn, Mn, Fe, Co, and Al in drinking water samples based on novel extraction technique using atomic absorption spectrometry in Turkey. According to their studies, the concentration of Cr in natural water was often very small. Based on results, the hexavalent form of chromium (Cr  $_{\rm VI}$ ) is ranged between 3.14 and 6.08 µgL<sup>-1</sup>. The mean of Cr levels in the waters of the area of Turkey was equal to4.40 µgL<sup>-1</sup>. Maximum tolerable limit of concentration of total chromium in drinking water was less than 50 µg L<sup>-1</sup> (Tlv of chromium by WHO  $> 50 \ \mu g \ L^{-1}$ ) [101]. The permissible value of Ni in drinking water was less than 20 µg L<sup>-1</sup>. The mean level of Ni in the water samples, in the area of Tokat, Turkey was equal to 3.82 µg L<sup>-1</sup> which was lower than standard references such as EPA, WHO. Also, according to their study, the drinking water sample from Turhal was the highest nickel value

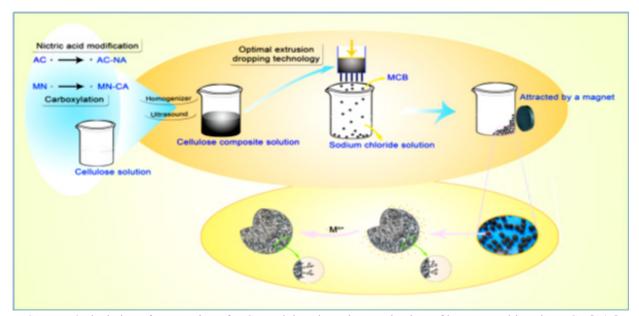


Scheme 2. Removal of heavy metal Ions by desorption process [139]

 $(5.35\pm0.28 \ \mu g \ L^{-1})$  and the city center of Tokat was the lowest  $(2.16\pm0.19 \ \mu gL^{-1})$  [101]. They showed, the amount of Cu was in the range of 4.44-7.43 µgL<sup>-1</sup>. In Erbaa, Turkey, Cu was substantially lower than the permissible limit defined by WHO (1.0  $\mu$ gL<sup>-1</sup>) in drinking waters [101]. In addition, mean level of concentration of Cu was equal to 6.01 µg L<sup>-1</sup>. Therefore, no contamination of Cu was there in the drinking water samples. The guideline value for zinc in drinking-water is given as 5.0 mg/l by WHO. The concentration of Zinc ranged from 4.16 to 8.44  $\mu$ g L<sup>-1</sup> in the drinking water samples. The mean level of zinc was obtained 6.12  $\mu$ g L<sup>-1</sup>, which was more than the permissible limit value  $(5.0 \text{ mg/L}^{-1})$  [101]. Many published methods were reported for determination and sample preparation of heavy metals in water and biological samples [Table 1-3]. Chromium speciation based on acetylcysteine (NAC) by dispersive liquidliquid bio-microextraction was achieved in human blood samples by shirkhanloo et al. By method, Ionic liquid (1-hexyl-3-methylimidazolium hexafluorophosphate) was rapidly injected into the blood samples containing Cr<sup>3+</sup>, which have already complexed by NAC at pH from 5 to 8. By optimizing, the linear range, limit of detection and preconcentration factor were achieved 0.03-4.4 µg  $L^{-1}$ , 0.005 µg  $L^{-1}$ , and 10 respectively [163,164]. Also, chromium speciation was reported by other

researcher [165-172]. Xiaogang Luo et al explained a novel sorbent for absorption heavy metals from waters by syntesis of MCB. The carboxyl decorated magnetite (Fe<sub>3</sub>O<sub>4</sub>) nanoparticles (MN-CA) were obtained by postmodifying the magnetite surface with citric acid. The nanoparticles of Fe<sub>3</sub>O<sub>4</sub> were added to 0.1 M citric acid under ultrasonic condition for 45 min, and the reaction was kept for 4 h at room temperature. A magnet was used to separate the MN-CA, and then the prepared MN-CA samples were rinsed thoroughly with acetone and double distilled water. The proposed method based on MN-CA followed by Scheme 3 for adsorption heavy metals from waters [145].

Naeemullaha et al showed, the effect of main parameters on extraction of vanadium in waters was evaluated by  $\mu$ S-SHS procedures [102]. For optimizing, Statistical assessment of  $\mu$ S-SHS method, Screening and optimization of experimental variables and selectivity of the  $\mu$ S-SHS method was studied and finally, the method was validated by CRM (SLRS-4 Riverine water samples). The optimization of experimental variables followed with screen out of the volume of SHS, pH, volume of Na<sub>2</sub>CO<sub>3</sub> and volume of H<sub>2</sub>SO<sub>4</sub> solution as a extraction efficiency with t-value (95 %) of 2.365. The simple, fast and switchable hydrophilicity microextraction (SHM) couple with ETAAS through micropipette tip



Scheme 3. depiction of preparation of MCB and the adsorption mechanism of heavy metal ions by MCB [145].

syringe system ( $\mu$ S-SHS) was used for separation and determination of vanadium in food and water samples. The  $\mu$ S-SHS method introduced an ideal sample preparation for vanadium extraction. The simple, fast and applied method based on SHM had good efficiency as compared with conventional methods for extraction of vanadium [102]. Also, the proposed method based on  $\mu$ S-SHS compared with published method and results showed high precision and accuracy results with low LOD and RSD (Table 1). Moreover, the graphical abstract of the  $\mu$ S-SHS was shown in Figure 3.

Many separation methods were used for determination of ultra-trace metals in different matrix, such as Liquid-liquid microextraction, dispersive liquid–liquid microextraction, solidphase microextraction, selective dispersive micro solid-phase extraction, traditional organic solvents, and switchable solvent, ultrasound assisted-dispersive-ionic liquid-micro-solid phase extraction and ultrasound assisted-dispersive solid-liquid multiple phase microextrbased on action [Table 1-3]. Kaiser, Zheng and Feng used a novel two-dimensional polymers crystals (TDPC) synthesis with nano size in waters through reaction between amine and anhydride monomers by surfactant monolayers. They showed high crystallinity polymers (CsP) with thickness (2 -3 nm) and an crystal size ( $3.5 \mu m^2$ ). The molecular structure of the materials, were characterized using X-ray scattering and TEM. The structure and TEM was shown in Figure 4. The formation of CsP is attributed to the pre-organization of monomers at interface phase (water- surfactant). Finally, micrometre-sized and few layer TDPC were grown. This material was used in different application such as removal heavy metals [156].

Rosillo-Lopez et al used carboxylated graphene nanoflakes (CX-GNF) and nanographene oxide (GO) for extracting of heavy metals such as,  $Fe^{2+}$ ,  $Cu^{2+}$ ,  $Fe^{3+}$ ,  $Cd^{2+}$  and  $Pb^{2+}$  in waters. The carboxylic acid groups (cx-GNF) have strong binding with heavy metal cations. CX-GNF with additional chemicalfunctionalization procedures can easily compete

**Table 1.** The proposed method based on  $\mu$ S-SHS compared with published method [102]

Sample prepration	Technique	SV	RSD%	LOD (µg.L <sup>-1</sup> )	EF/PF	References	
SPE	ICP-OES	10	3.4	0.06	45	68	
CPE approach	Spectrometric		••••	1.4		70	
CPE	ETAAS	50		0.042	125	71	
μS-SHS	ETASS	10	2.2	0.0075	120	76	

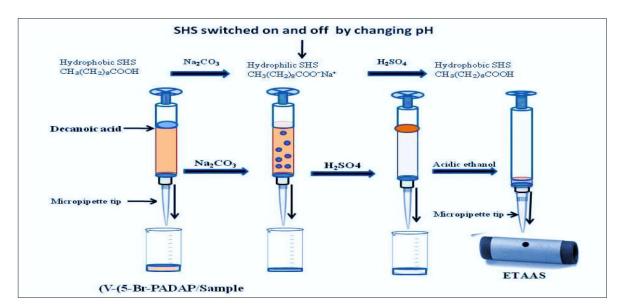


Fig. 3. The graphical abstract of the  $\mu$ S-SHS [102].

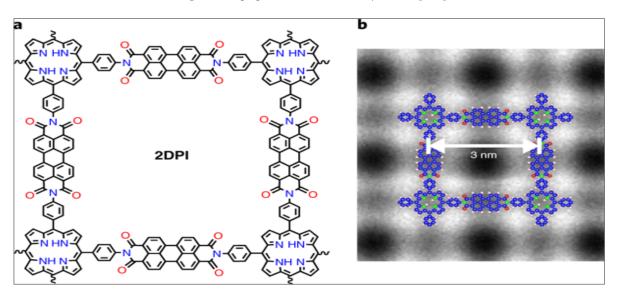


Fig. 4. The structurea and b) TEM of two-dimensional polymers crystals (TDPC)[156]

with other carbon materials such as G and GO for heavy-metal extraction (Fig. 5). Furthermore, the cx-GNF material extracted Pb<sup>2+</sup> ions in the presence of excess Ca<sup>2+</sup> or Mg<sup>2+</sup> cations which were already existed in environmental matrix. So, the CX-GNF material showed the highest potential for extraction of heavy-metal from water as compared to other graphene materials [157]. Zhao et al used multithroughput dynamic microwave-assisted leaching (MDMAL) for determination of Cu, Mn, Zn and Pb in soil . This method combines the microwave and dynamic leaching technique for five samples within 15 min (Fig. 6). HNO<sub>3</sub> aqueous solution was

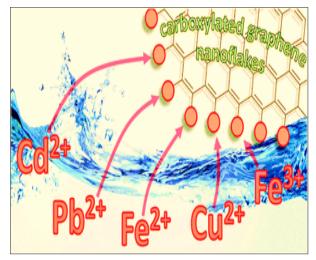


Fig. 5. Heavy metal extraction by cx-GNF.

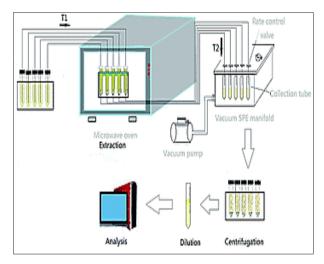
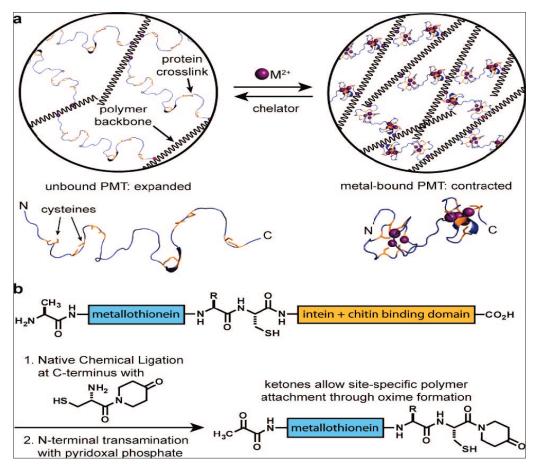


Fig. 6. Heavy metal extraction by MDMAL

employed as leachant at a flow rate of 1.5 mL min<sup>-1</sup>. The leaching condition were above 95% for Cu, Mn, Zn, Pb and lower for Cr, Cd, Ni and Co, compared to the conventional method. The mean of RSD of intraand inter-day of Cu, Mn, Zn, and Pb were 4% and 6%, respectively. The proposed method was more rapid than the conventional digestion method and could be an alternative approach for the analysis of heavy metals in complex solid samples [158].

Aaron et al, showed that, the crosslinking of polymer chains based on the N- and C- of proteins  $(CPC \rightarrow N/C-P)$  was used as a key of the proteins which was captured trace amounts of heavy metals such as cadmium in the presence of innocuous ions in contaminated water sample. Protein-cross-linked hydrogels introduced for sequestration of heavy metal ions in water samples. As Figure 7, the Pea metallothioneins (PMTs) can capture toxic metal ions by condensing to form binding pockets. The synthetic route to access these materials involves the introduction of ketones on both protein termini. These groups are then used to cross-link alkoxyamine-substituted polymers through oxime formation [159]. Shirkhanloo et al used NH<sub>2</sub>-UVM<sub>7</sub> based on US-D-µ-SPE method for speciation of manganese in water samples. The batch adsorption



**Fig. 7.** Pea metallothioneins (PMTs) can capture toxic metal ions a) Condensing to form binding pockets b) Ketones used to cross-link alkoxyamine-substituted polymers [159]

capacity of NH<sub>2</sub>-UVM<sub>7</sub> for Mn(II) and Mn(VII) ions was found to be 1733  $\mu$ mol g<sup>-1</sup> and 570  $\mu$ mol g<sup>-1</sup>, respectively. In optimized parameters, LOD (3Sb/m), and LOQ (10Sb/m) were 0.007  $\mu$ g L<sup>-1</sup> and  $0.03 \ \mu g \ L^{-1}$  for Mn(II). The mean preconcentration factors for Mn(II) and Mn(VII) were calculated as 102.3 and 98.8 with RSD about 2.8%, respectively (Fig. 8). The regression equations for the calibration curves of Mn(II) and Mn(VII) were A=9.8438 C+0.0053 (R2=0.9997), and A=9.6922 C+0.0045 (R2=0.9995), respectively [160]. In addition, Shirkhanloo et al used NG-COOH for speciation of trace mercury in waters by US-D-IL- µ-SPE. Under optimized conditions, the linear range, LOD and PF/EF were obtained 0.03–6.3  $\mu$ g L<sup>-1</sup>, 0.0098  $\mu$ g L<sup>-1</sup>and 10.4 for caprine blood samples, respectively. The developed method was successfully applied to natural water and human/caprine blood samples. In order to validate the method described, two certified standard reference materials, NIST-SRM 995c (mercury species in caprine blood) and NIST-SRM1641e (total mercury inwater), were analyzed by US-D-IL- μ-SPE [161]. They used NH2-UVM7 based on USA-DSLMPME for speciation of trace amounts arsenic (III and V) in waters and human urine samples. The linear range, LOQ  $(3\sigma)$ , RSD% and EF for As(V)were obtained 0.02-1.65 µg  $L^{-1}$ , 11 ng  $L^{-1}$ , 4.3% and 100.5. The recoveries (95-102%) was achieved by certified reference material in urine (NIST-SRM). The mean of LOD

was obtained 3.3 ng  $L^{-1}$  and 2.7 ng  $L^{-1}$  for human urine and standard samples, respectively (intra-day, Mean of LOD = 3)[162].

## 3.1. Chemistry techniques for heavy metals

Also, many chemistry techniques were reported in published papers for separation, extraction, preconcentration and speciation metals in different matrixes (Table 2). Recently, the liquid-liquid extraction methods (LLE) were used for separation and determination heavy metals from water and human biological samples. Some of LLE techniques such as, ultrasonic-thermostatic-assisted cloud point extraction procedure (UTA-CPE-FAAS), aqueous solvent-based dispersive liquid-liquid microextraction (AS-DLLME), cloud point extraction - slotted quartz tube-flame atomic absorption spectrometry (CPE-SQT-FAAS), surfactant-assisted dispersive liquid-liquid micro-extraction (SA-DLLME-FAAS), switchable liquid-liquid microextraction (SLLME-SQT-FAAS), surfactant-Based Dispersive Liquid-Liquid Microextraction based on FAAS(SB-DLLME-FAAS), vortex-assisted DLLME based on voltammetric (VA-DLLME-VM), microwaveassisted extraction - inductively coupled plasmaoptical emission spectrometry (MAE-ICP-OES), Ionic liquid-based microwave-assisted dispersive liquid-liquid microextraction (IL-based MA-DLLME), Ionic liquid-based dispersive liquidliquid microextraction (IL-DLLME), Ionic liquid

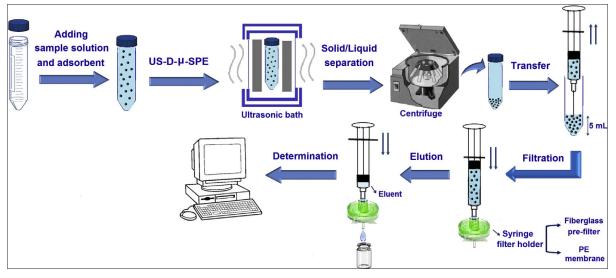


Fig. 8. Procedure of Mn speciation based on NH<sub>2</sub>-UVM<sub>7</sub> by US-D-µ-SPE procedure [160].

Sample	Metals	Method	Solvent /Ragent	EF/PF	LOD $\mu g L^{-1}$	<b>Ref.</b> [173]
Wastewater	Cd	DLLME/SQT-FAAS	DPC	93.3	0.5	
Milk	V(V) and Mo(VI)	UTA-CPE-FAAS	Nile blue A -	145 and 115	0.86 and 1.55	[174]
Vegetables			PONPE			
Foodstuffs			DDC	(0)	0.0007	F 1
Parsley	Cd	CPE-SQT-FAAS	DPC	60	0.0007	[175]
Тар	Zn	SA-DLLME-FAAS	PAR		0.5	[176]
River						
Well water	G		DDC	0656	0.07	[177]
Wastewater	Co	DLLME/SQT-FAAS	DPC	86.56	0.97	[177]
Tea	Со	SLLME-SQT-FAAS	Schiff base ligand	107.7	3.1	[178]
vitamin B12	C (II)		Trite X 114	50	1 (1	[170]
Food	Cu(II)	SB-DLLME-FAAS	Triton X-114	50	1.61	[179]
Water Samples Milk	Cd(II), Cu(II), Pb(II)	DLLME/FAAS	TDES		0.38-0.42	[180]
Chocolate	Ni	<b>PV-IS-DLLME</b>	APDC	17	100.0	[181]
Urine	Hg	VA-DLLME-VM	AuNPS-PE		1.1 - 1.3	[182]
human blood	Cr(III and VI)	CP-DILLME-ET-AAS	IICDET	25.2	0.0054	[183]
Human serum	Cd	DLLME-ETAAS	TSILs	10.2	0.005	[184]
Human blood	Cr(III and VI)	IL-DLLBME-ETAAS	NAC	10	0.005	[185]
Biodiesel	Na, K, Ca and Mg	<b>RP-DLLME/FAAS</b>	Iisopropanol,HNO <sub>3</sub> ,		0.006- 0.026	]186]
Water	Ta (I), Ta (III)	IL DLLME-ET-AAS	[C6MIm][PF6]	100	3.3 ng L <sup>-1</sup>	[187]
Cosmetic	Hg (II), MeHg+,EtHg+	HPLC-ICPMS	[C6MIm][PF6]	760, 115, 235	1.3 ng L <sup>-1</sup>	[188]
Wines	As (III)	DLLME-ETAAS	[C8MIm][PF6]		$5.0 \text{ ng } \text{L}^{-1}$	[189]
Water	Cu(II)	DLLME-FAAS	[C4MIm][PF6] or	54	$3 300 \text{ ng } \text{L}^{-1}$	[190]
			[C16C4Im][Br]		8	
Water and	Rh (III)	US-IL DLLME-FAAS	[C8MIm][NTf2]		0.37	[191]
leaves						
Water	$V\left(IV\right)$ and $V\left(V\right)$	TC-IL DLLME-ETAAS	[C4MIm][PF6]		0.0049	[192]
Tea, Soda	Se (IV)	USA-IL-DLLME-	[C6MIm][NTf2]	150	12	[193]
Beer		ETAAS				
Milk						
Water	Au, Ag	MRTILs- DLLME- ETAAS	$[C_5(MIM)_2][NTf_2]_2$	245 and 240	3.2 and 3.7	[194]
Food	Zn, P, Cd, Pb, Fe, Mn,	MAE -ICP-OES	MAE		0.01 mg kg <sup>-1</sup>	[195]
	Mg, Cu, Ca, Al, Na, K				$-7.8 \text{ mg kg}^{-1}$	
Water	Pd and Pt	CPE-ICP OES	MBT		0.525 and 0.752	[196]
Water serum	Cu(II)	IL-mE-DLLME-CPE- ETAAS	Oxine-IL	70	0.132	[197]

Table 2 Solvent extraction for determination and separation of heavy metals in different matrix [173-197]

EF: Enrichment Factor

UTA-CPE-FAAS: ultrasonic-thermostatic-assisted cloud point extraction procedure

AS-DLLME: Aqueous solvent-based dispersive liquid-liquid microextraction

DPC: Diphenylcarbazone

Nile blue A: 9-(diethylamino)benzo[a]phenoxazin-5-ylidene]azanium; sulfate-PONPE (polyoxyethylene-nonylphenyl ether

ETAAS: Electrothermal atomic absorption spectrometry

FAAS: Flame atomic absorption spectrometry

SQT-FAAS: Slotted quartz tube-flame atomic absorption spectrometry

CPE-SQT-FAAS: Cloud point extraction - slotted quartz tube-flame atomic absorption

spectrometry

DPC: Diphenylcarbazone

SA-DLLME-FAAS : Surfactant-assisted dispersive liquid-liquid micro-extraction

PAR: 4-(2-pyridylazo) resorcinol

SLLME-SQT-FAAS: Switchable liquid-liquid microextraction

SB-DLLME-FAAS: Surfactant-Based Dispersive Liquid-Liquid Microextraction/FAAS Triton X-114: Surfactant

TDES: Ternary deep eutectic solvent

APDC: ammonium pyrrolidine dithiocarbamate

VA-DLLME-VM: Vortex-assisted DLLME/voltammetric

AuNPS-PE: gold nanoparticle-modified screen-printed electrodes

MAE-ICP-OES: microwave-assisted extraction - inductively coupled plasma-optical

emission spectrometry

MBT: 2-mercaptobenzothiazole

Oxime-IL: 8-hydroxyquinoline-IL

UA-DLLME: Ultrasound-assisted dispersive liquid-liquid microextraction

UAE- IL-DLLME: Ultrasonic-assisted ionic liquid based dispersive liquid-liquid microextraction

USA-DLLME: Ultrasound-assisted dispersive liquid-liquid microextraction

USA-IL-DLLME: Ultrasound assisted ionic liquid dispersive liquid-liquid microextraction

ultrasound assisted dispersive liquid-liquid microextraction (IL-UA-DLLME), In-situ ionic liquid dispersive liquid-liquid microextraction(in-situ IL-DLLME), microwave-assisted extraction and dispersive liquid-liquid microextraction (MAE-DLLME), magnetic ionic liquid-based dispersive liquid-liquid microextraction) MIL-based DLLME), magnetic stirring-assisted dispersive liquid-liquid microextraction (MSA-DLLME), magnetic stirrer induced dispersive ionic-liquid microextraction (MS-IL-DLLME), microwave-assisted dispersive liquid-liquid microextraction(MWA-DLLME), ionic liquid dispersive microextraction (IL-DLLME), surfactant assisted dispersive liquidliquid microextraction (SA-DLLME), switchable solvent-based dispersive liquid-liquid microex-

Table 3. Different sorbents for extraction of heav	y metals in human and environmental samples [1	198-218]
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Sample	Method	Sorbent	Heavy metals	LOD (ng $L^{-1}$ )	EF/PF	AC(mg g <sup>-1</sup> )	Ref.
Serum Blood	USA-D-µSPE-ETAAS	NH <sub>2</sub> -UVM <sub>7</sub>	Cd	2.0	25	108.6	[198]
Waters	MCD-µSPE-ETAAS	GSH	Pb, Cd, Cr	8.9, 4.0, 10	17, 18,17	146.3, 132.5, 125.8	[199]
Waters	USA-CP-MSPE	AFSA	Pb	10.0	102		[200]
River							
Urine	USA-DSL-MPME	NH <sub>2</sub> -UVM <sub>7</sub>	As $_{\rm III}$ , As $_{\rm V}$	1.1	100.5		[201]
Water							
Blood	US-D-IL- µSPE-AT-	NG-COOH	Hg, MHg, EHg	9.8	10.4	153.7, 125.8, 125.8	[202]
Water	FAAS						
Water	US-D- µSPE	NH <sub>2</sub> -UVM <sub>7</sub>	Mn $_{_{\rm II}}$ , Mn $_{_{\rm VII}}$	7.0, 8.0	102.3, 98.8	1733 and 570	[203]
Tap W		G 0.1 75		• • •		μmol.g <sup>-1</sup>	500.43
	USA-D- µSPE	GONPs	Al	20.0	25	187.5	[204]
Water	SPE-FAAS	CdSNPs	Pb, Cu			200, 166.7	[205]
Drinking		~					
Wastewater	FTIR	G+MGO	Pb <sup>2+</sup> , d <sup>2+</sup> ,Cu <sup>2+</sup>			358.96, 388.4, 169.8	[206]
Waters	SPE-FAAS	GO	Pb, Ni	1400-2100	95-102.5	195, 178	[207]
Waters	ET-AAS	GO-NH <sub>2</sub>	Pb (II)	9.4	100	479	[208]
serum	H-US-D-µ-SPE	G-COOH	As species	2.1	53	125.4	[209]
urine							
River water	CHD-µSPE	GO	Pb, Cd, Cr	110, 15, 38	15		[210]
Waters	SPE-ETAAS	MnO <sub>2</sub> / CNT	PB, Cd	4.4, 1.5	100		[211]
Waters	DµSPE-TRXRFS	MWCNT	Cr <sub>(VI)</sub>	3000	66	154.3	[212]
Urine							
Soils	SPE-ICPMS	Ti- MSN	Cu	3.9	10		[213]
Waters							
Blood	M-SPµE-FAAS	Modified	Cd	150	10		[214]
		carbon cloth					
FOOD	SPE-FAAS	Dowex	Pb	130	250		[215]
A		Marathon C	7	200	220		[217]
Agricultur	SPE-FAAS	F-MOF	Zn	200	238		[216]
Foods	SPE-FAAS	B2O3/TiO2	Cd	1440	50	172.5	[217]
Waters	D-µSPE-ETAAS	NH <sub>2</sub> -SiO <sub>2</sub> -GO	Pb	9.4	100		[218]

AC: Absorption Capacity

EF/PF: Enrichment factor/preconcentration factor

ET-AAS: Electro-thermal atomic absorption spectrometry

FAAS: Flame atomic absorption spectrometry

GSH: Graphene-silica hybrid

USA-D-µSPE: Ultrasound assisted-dispersive-micro solid phase extraction MCD-µSPE: Moderate centrifugation-assisted dispersive micro solid phase extraction

USA-CP-MSPE: Ultrasound-assisted cloud point-micro solid phase extraction AFSA: Amine functionalized silica aerogel

USA-DSL-MPME: Ultrasound assisted-dispersive solid-liquid multiple phase

microextraction

US-D-IL-µ-SPE: ultrasound assisted-dispersive-ionic liquid-micro-solid phase extraction

NG: Nanographene

CHD-µSPE: Coagulating homogenous dispersive micro-solid phase extraction CdSPNs: Cadmium Sulfide Nanoparticles

TRXRFS: Total reflection X-ray fluorescence

GONPs: Graphene oxide nano-particles

M-SPµE: Miniaturized solid phase microextraction

SPE-FAAS: Solid phase extraction-flame atomic absorption spectrometry

GO-NH, Aminosilanized graphene oxide

traction (S-DLLME), temperature-assisted ionic liquid-based dispersive liquid–liquid Microextraction (TA-IL-DLLME), ultrasound-assisted dispersive liquid-liquid microextraction (UA-DLLME), ultrasonic-assisted ionic liquid based dispersive liquid–liquid microextraction (UAE- IL-DLLME), ultrasound-assisted dispersive liquid-liquid microextraction (USA-DLLME), ultrasound assisted ionic liquid dispersive liquid-liquid microextraction (USA-IL-DLLME) was used for heavy metal extraction in different matrixes (Table 2) [173-197]. Many researchers were reported other nanomaterial or sorbents for extraction metals and heavy metals from water, wastewaters, air pollution and human biological matrix which were shown in Table 3.

## 3.2. Nano particles for extraction heavy metals

The affinity of Hg towards Au was generally

achieved by AuHg, AuHg3, and Au3Hg [219]. Lisha et al. reported the removal of Hg (II) with nanoparticles of Au (AuNPs) coated on aluminum [220]. NaBH4 helped to reduce Hg (II) to Hg (0) and removed by AuNPs with capacity of 4.0 g g<sup>-1</sup>. Also, Jiménez et al. was used citrate-coated AuNPs for extraction mercury in water [221]. Absorption mercury with AuNPs was shown in Figure 9.

Magnetite  $(Fe_3O_4)$  as nanoadsorbent was used for heavy metal removal as environmental friendliness.  $Fe_3O_4$  could be easily separated from liquid phase.  $Fe_3O_4$  was used for heavy metals treatment in waters [222-224]. Giraldo et al. synthesized magnetite nanoparticles by using a coprecipitation method and were used for extracting of Pb (II) and Mn (II) in different samples [225]. In Figure 10, the removal of chromium was shown by chitosan–magnetite nanocomposite.

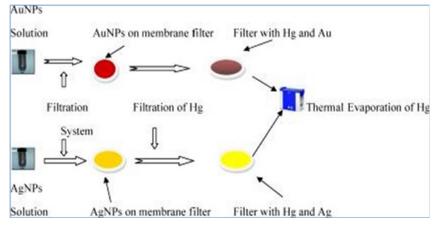


Fig. 9. The absorption mercury by AuNPs

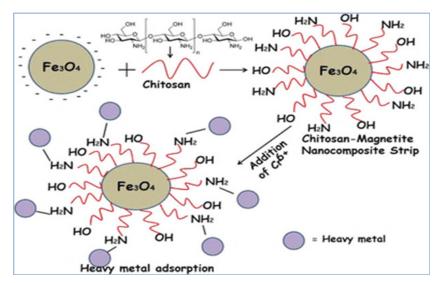


Fig. 10. The removal of chromium by chitosan-magnetite nanocomposite strip

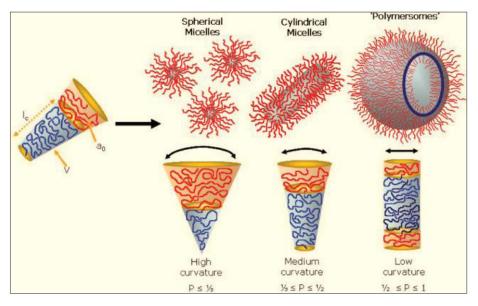
Magnetite particles are modified with groups such as, MPs-NH2 [226], MPs–COOH [227], MPs– SH [228]. The core-shell structure of PI-b-PEG diblock copolymer encapsulated with iron oxide nanoparticles [229]. PI-b-PEG diblock copolymer was used for removal heavy metals from waters [230]. The nanostructures of amphiphilic diblock copolymers due to the inherent curvature of the polymer were shown in Figure 11 which was used for extraction lead and other heavy metals from different matrixes.

Zero valent iron as nanoparticles of is a Fe (0)and ferric oxide coating (nZVI) which was used for extraction heavy metals from different matrix (Figure 3) [231]. Also, nZVI as a novel adsorbent can be removed heavy metals (Fig. 12), such as mercury, chromium, copper, nickel, and cadmium from waters. [232-235]

The silica nanoparticles based on ligand of  $\beta$ -ketoenol-pyridine-furan (as a selective adsorbent was used for metal removal from waters (Pb,Cd,Zn,Cu) (Scheme 4). The engineered sorbent had high adsorption capacity for capturing toxic heavy metal in environmental samples [236-239].

## 4. Conclusions

In this review paper, constructive issues from novel published papers were selected for studying



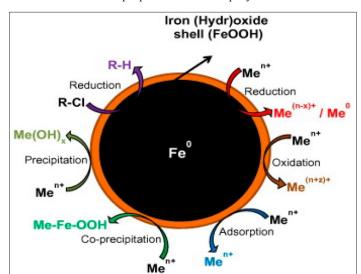
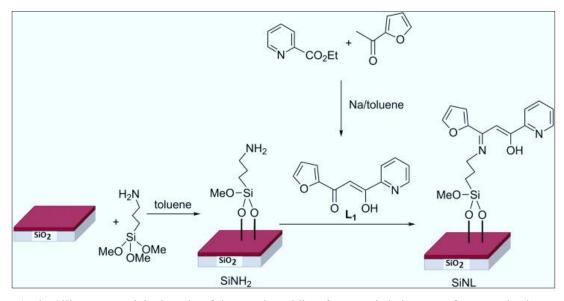


Fig. 11. The nanostructures of amphiphilic diblock copolymers for extraction lead ions

Fig. 12. The core-shell structure of nZVI used for extraction of metals [231]



Scheme 4. The Silica nanoparticles based on  $\beta$ -ketoenol-pyridine-furan as chelating agent for extraction heavy metals in liquid phase

of separation, preconcentration, and extraction of heavy metals from water, waste water, vegetable and human biologic samples. Recently, the SPE methods were coupled with spectrometry techniques such as atomic absorption spectrometry (AAS) based on various adsorbents included graphene, graphene oxide, activated carbon, carbon nanotubes (CNTs), zeolites, MOF, magnetic nanoparticles, carbon quantum dots (CQDs)a nd MSN which was used for ultra-trace determination of heavy metals in different matrixes. In addition, the liquid-liquid micro extraction techniques coupled with ET-AAS, F-AAS, ATF-AAS, SQT-FAAS, ICP-XRF, CV-AAS, ICP, ICP-AES, ICP-MS, ICP-OES, HPLC, voltammetry, UV-VIS, and FTIR were presented in this study by a lot of researchers. The proposed review showed us, the novel techniques based on nanotechnology had been developed in SPE and LLE methods since 2010 by authors. Also, SPE and solvent extraction based on different adsorbents and instrumental techniques were extended in various sciences such as, bio chemistry, nanotechnology, environmental analytical chemistry and human analysis during last decade.

## 5. References

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